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NEWS 18 Aug 01 NIS has been reloaded and enhanced  
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now available on STN  
NEWS 20 Aug 14 IFIPAT, IFICDB, and IFIUDB have been reloaded  
NEWS 21 Aug 14 The MEDLINE file segment of TOXCENTER has been reloaded  
NEWS 22 Aug 20 Sequence searching in REGISTRY enhanced  
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NEWS 24 Sep 1 Experimental properties added to the REGISTRY file  
NEWS 25 Sep 10 Indexing added to some pre-1967 records in CA/CAPLUS  
NEWS 26 Sep 10 CA Section Thesaurus available in CAPLUS and CA  
NEWS 27 Oct 01 CASREACT Enriched with Reactions from 1907 to 1985  
NEWS 28 Oct 21 EVENTLINE has been reloaded  
  
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L4 4 L3 (S) RADICAL(W) MEDIAT?

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L1 ANSWER 1 OF 4 BIOSIS COPYRIGHT 2002 BIOLOGICAL ABSTRACTS INC.

AN 1999:119009 BIOSIS

DN PREV199903119009

TI Increased DNA oxidation and decreased levels of repair products in Alzheimer's disease ventricular CSF.

AV Lovell, Mark A.; Gambita, S. Prasad; Markesbery, William R. 11

CA 111 Sanders-Brown Build., Univ. Kentucky, Lexington, KY 40536-0131

DA

SO Journal of Neurochemistry, (Feb., 1999) Vol. 72, No. 2, pp. 771-776.

ISSN: 0022-3042.

BT Article

LA English

L5 ANSWER 2 OF 4 MEDLINE

AN 1999127943 MEDLINE

DN 99117943 PubMed ID: 9939752

TI Increased DNA oxidation and decreased levels of repair products in Alzheimer's disease ventricular CSF.

AU Lovell M A; Gabbita S P; Markesbery W R

CS Sanders-Brown Center on Aging, and Department of Chemistry, University of Kentucky, Lexington 40536-0230, USA.

NC 1901-AG05118 (NIA)

SP 11-AG05144 (NIA)

SO JOURNAL OF NEUROCHEMISTRY, (1999 Feb) 72 (2) 771-6.

Journal code: 2985190R. ISSN: 0022-3042.

CY United States

DT Journal; Article; (JOURNAL ARTICLE)

LA English

FS Priority Journals

EM 122902

ED Entered STN: 19991223

Last Updated on STN: 19991223

Entered Medline: 19991211

L5 ANSWER 3 OF 4 EMBASE COPYRIGHT 2002 ELSEVIER SCI. B.V.

AN 1999042104 EMBASE

TI Increased DNA oxidation and decreased levels of repair products in Alzheimer's disease ventricular CSF.

AU Lovell M.A.; Gabbita S.P.; Markesbery W.R.

CS Dr. W.R. Markesbery, 101 Sanders-Brown Building, University of Kentucky, Lexington, KY 40536-0230, United States

SO Journal of Neurochemistry, 1999) 72/2 (771-776).

Refs: 44

ISSN: 1931-3042 CODEN: JONFA

CY United States

DT Journal; Article

FS 005 General Pathology and Pathological Anatomy

006 Neurology and Neurosurgery

LA English

SL English

L5 ANSWER 4 OF 4 LIFESCI COPYRIGHT 2002 CSA

AN 1999091917 LIFESCI

TI Increased DNA Oxidation and Decreased Levels of Repair Products in Alzheimer's Disease Ventricular CSF

AU Lovell, M.A.; Gabbita, S.P.; Markesbery, W.R.\*

CS 101 Sanders-Brown Building, University of Kentucky, Lexington, KY 40536-0230, USA

SO Journal of Neurochemistry [J. Neurochem.], (19990200) vol. 72, no. 2, pp. 771-776.

ISSN: 0022-3042.

DT Journal

FS N3

LA English

SL English

AS

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AN 1999:526234 CAPLUS

DN 131:309477

TI Fluorescent labelling of closely-spaced aldehydes induced in  
DNA by bleomycin-Fe(III)

AU Chakrabarti, S.; Mahmood, A.; Makrigiorgos, G. M.

CS Joint Center for Radiation Therapy and Dana Farber Cancer Institute,  
Harvard Medical School, Boston, MA, 02215, USASO International Journal of Radiation Biology (1999), 75(8), 1055-1065  
CODEN: IJRBEE; ISSN: 1955-3002

PB Taylor &amp; Francis Ltd.

DT Journal

LA English

AB The purpose of this study was to test the ability of two novel fluorescent reagents fluorescent aldehyde-reactive probe (FARP) and FARPhe, to label aldehyde-contg. sites (principally apurinic sites) generated in DNA by the radiomimetic drug bleomycin, and to use fluorescent energy transfer from FARPhe (donor) to FARP (acceptor) to quantitate such closely-spaced sites. FARPhe, 7-hydroxycoumarin-3-carboxylic acid (((((amino-oxymethyl) carbonyl) hydrazino) carbonyl) ethyl) amide) was synthesized with a protocol similar to the one recently reported for FARP (a fluorescein-based probe). Both FARPhe and FARP form stable covalent bonds with the open-chain aldehydes generated upon acidic depurination of DNA. Plasmid DNA exposed to bleomycin-Fe(III)-ascorbate undergoes extensive strand breakage, and upon subsequent reaction with FARPhe and/or FARP it becomes fluorescently labeled, indicating the generation of aldehyde-contg. sites. The binding of the probes to calf thymus or plasmid DNA results in significant fluorescent energy transfer among closely-spaced fluorophores, as revealed by the fluorescence increase following digestion of fluorescently labeled samples with nuclease P1. The fluorescence quenching is most evident when both FARPhe and FARP are used simultaneously to trap aldehyde sites. When single-stranded oligonucleotides engineered to contain either one or two closely spaced bleomycin binding sites are exposed to bleomycin and then fluorescently labeled, the oligonucleotides demonstrate significantly increased fluorescent energy transfer with two binding sites indicating a dependence of aldehyde site generation and clustering on the local sequence of a single strand. In conclusion, a new detection method for DNA damage induced by bleomycin following fluorescent labeling of aldehyde group-contg. sites (FLAGS) and their clustering via fluorescent energy transfer is demonstrated. The method is applicable to any form of DNA. This work may lead to a general approach for the quantification of multiply damaged sites in DNA, a subset of DNA lesions that may have major biological significance.

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L3 ANSWER 8 OF 16 MEDLINE DUPLICATE 3  
 AN 97105898 MEDLINE  
 DN 97105898 PubMed ID: 8948646  
 TI Chemical methods of DNA and RNA fluorescent labeling.  
 AU Prudnikov D; Mirzakekov A  
 CS Engelhardt Institute of Molecular Biology, Moscow, Russia.  
 SO NUCLEIC ACIDS RESEARCH, (1996 Nov 15) 24 (22) 4535-42.  
 Journal code: CBL; 0411011. ISSN: 0305-1048.  
 CY ENGLAND: United Kingdom  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 EM 199701  
 ED Entered STN: 19970219  
 Last Updated on STN: 19970219  
 Entered Medline: 19970117  
 AB Several procedures have been described for fluorescent labeling of DNA and RNA. They are based on the introduction of aldehyde groups by partial depurination of DNA or oxidation of the 3'-terminal ribonucleoside in RNA by sodium periodate. Fluorescent labels with an **attached** hydrazine group are efficiently coupled with the aldehyde groups and the hydrazone bonds are stabilized by reduction with sodium cyanoborohydride. Alternatively, DNA can be quantitatively split at the depurinated sites with ethylenediamine. The aldimine bond between the aldehyde group in depurinated DNA or oxidized RNA and ethylenediamine is stabilized by reduction with sodium cyanoborohydride and the primary amine group introduced at these sites is used for **attachment** of isothiocyanate or succinimide derivatives of fluorescent dyes. The fluorescent DNA labeling can be carried out either in solution or on a reverse phase **column**. These procedures provide simple, inexpensive methods of multiple **DNA labeling** and of introducing one fluorescent dye molecule per RNA, as well as quantitative DNA fragmentation and incorporation of one label per fragment. These methods of fluorophore **attachment** were shown to be efficient for use in the hybridization of labeled RNA, DNA and DNA fragments with oligonucleotide microchips.

L3 ANSWER 10 OF 16 MEDLINE DUPLICATE 4  
 AN 94057384 MEDLINE  
 DN 94057384 PubMed ID: 8238885  
 TI Biotinylation of **DNA** on membrane **supports**: a procedure  
 for preparation and easy control of **labeling** of nonradioactive  
 single-stranded **nucleic** acid probes.  
 AU Eidenko V V  
 CS Department of Immunology, Institute of Transplantology and Artificial  
 Organs, Moscow, Russia.  
 SO ANALYTICAL BIOCHEMISTRY, (1993 Aug 15) 213 (1) 75-8.  
 Journal code: 4NK; 0370535. ISSN: 0003-2697.  
 CY United States  
 DT Journal; Article; (JOURNAL ARTICLE)  
 LA English  
 FS Priority Journals  
 EM 199312  
 ED Entered STN: 19940117  
 Last Updated on STN: 19940117  
 Entered Medline: 19931203  
 AB We have used M13 single-stranded DNA **bound** by uv to small pieces  
 of nylon membrane for the synthesis of biotinylated single-stranded DNA  
 probes. The labeling method requires a large fragment of DNA polymerase I  
 and random hexanucleotides. There is no need for previous linearization of  
 the template. The clean probe is removed from the membrane by a single  
 wash step. The synthesized probe is completely free of unincorporated  
 precursors. This makes possible the easy control of the reaction of  
 incorporation of biotinylated analogues into the probe by simple staining  
 on the filter, thus allowing evaluation of the efficiency of labeling. The  
 DNA membrane can be stored for reuse. With the procedure described it is  
 possible to biotinylate many DNA fragments in parallel, simultaneously  
 controlling the efficiency of labeling in a time- and cost-saving manner.